

Mouse HV-IgM/IgG (Hantaan Virus(HV) IgM/IgG) ELISA Kit

(Do not mix reagents of different batches and different product numbers in the kit, otherwise the kit will not work properly)

Catalogue No.: EM2185

Size: 96T

Reactivity: Mouse

Application: This immunoassay kit allows for the qualitative determination of HV-IgM/IgG in Mouse serum .

Storage: 2-8°C

Expiry Date: see kit label

Principle: Indirect

NOTE: FOR RESEARCH USE ONLY.

Kit Components

Item	Specifications(96T)	Storage
Micro ELISA Plate(Dismountable)	12 × 8	2-8°C/-20°C
HV-IgM/IgG Positive Control	1ml×1	2-8°C
HV-IgM/IgG negative Control	1ml×1	2-8°C
sample dilution buffer	12ml×1	2-8°C
HRP- Anti-Mouse IgM/IgG Antibody	12ml×1	2-8°C (Avoid Direct Light)
TMB substrate A	6ml×1	2-8°C (Avoid Direct Light)
TMB substrate B	6ml×1	2-8°C (Avoid Direct Light)
Stop solution	6ml×1	2-8°C
Wash buffer (20X)	50ml×1	2-8°C
Plate Sealer	3pieces	
Product Description	1 copy	

Principle of the Assay

This kit was based on indirect enzyme-linked immune-sorbent assay technology. HV-Ag was pre-coated onto 96-well plates. The test samples were added to the wells, unbound conjugates were washed away with wash buffer. Then added HRP- Anti-Mouse IgM/IgG Antibody, if there were any HV-IgM/IgG in the samples, it would form a HV-Ag- HV-IgM/IgG- HRP- Anti-Mouse IgM/IgG Antibody complex. The substrate TMB was added for color development, and the presence or absence of HV-IgM/IgG antibody was determined according to OD value after colorimetry on the enzyme label.

Precautions for Use

1. To inspect the validity of experiment operation and the appropriateness of sample dilution proportion, pilot experiment using standards and a small number of samples is recommended.
2. After opening and before using, keep plate dry.
3. Before using the kit, spin tubes and bring down all components to the bottom of tubes.
4. Storage TMB reagents avoid light.
5. Washing process is very important, not fully wash easily cause a false positive and high background.
6. Duplicate well assay is recommended for both standard and sample testing.
7. Don't let microplate dry at the assay, for dry plate will inactivate active components on plate.
8. Don't reuse tips and tubes to avoid cross contamination.
9. Please do not mix the reagents in different kits of our company. Do not mix reagents from other manufacturers.
10. To ensure accurate results, proper adhesion of plate sealers during incubation steps is necessary.

Material Required But Not Supplied

1. Microplate reader (wavelength: 450nm)
2. 37°C incubator
3. Automated plate washer
4. Precision single and multi-channel pipette and disposable tips
5. Clean tubes and Eppendorf tubes
6. Deionized or distilled water

Manual Washing

Discard the solution in the plate without touching the side walls. Clap the plate on absorbent filter papers or other absorbent material. Fill each well completely with 350ul wash buffer and soak for 1 to 2 minutes, then aspirate contents from the plate, and clap the plate on absorbent filter papers or other absorbent material. Repeat this procedure two more times for a total of THREE washes.

Automated Washing

Aspirate all wells, then wash plate THREE times with 350ul wash buffer. After the final wash, invert plate, and clap the plate on absorbent filter papers or other absorbent material. It is recommended that the washer be set for a soaking time of 1 minute.

Sample Collection and Storage

Isolate the test samples soon after collecting, then, analyze immediately (within 2 hours). Or aliquot and store at -20°C for long term. Avoid multiple freeze-thaw cycles.

- Serum: Coagulate the serum at room temperature (about 1 hours). Centrifuge at approximately 1000 × g for 15 min. Analyze the serum immediately or aliquot and store at -20°C.

Note: Samples to be used within 5 days may be stored at 2-8°C, otherwise samples must be stored at -20°C (≤1 month) or -80°C (≤2 months) to avoid loss of bioactivity and contamination. Hemolyzed samples are not suitable for use in this assay.

Wash Buffer Preparation:

Dilute 50mL of Concentrated Wash Buffer into 1000 mL of Wash Buffer with deionized or distilled water.

Assay Procedure

Remove the kit from the refrigerated environment and let it equilibrate at room temperature for 30 minutes before use

1. Wash plate 2 times before adding sample and control wells.
2. Label the sample wells, 3 Negative Controls, 2 Positive Controls and 1 blank well. Add 100 µL control serum to each well, except blank well.
3. Add 100µL of sample dilution to each of the other reaction Wells, and add 10µL of the test specimen in order. The color change effect is better when the sample is added and mixed by blowing.
4. Seal the plate with a cover and incubate at 37°C for 30 min.
5. Remove the cover, and wash plate 5 times with Wash buffer.
6. Add 100 µL HRP- Anti-Mouse IgM/IgG antibody to each well, except blank well
7. Seal the plate with a cover and incubate at 37°C for 30 min.
8. Remove the cover, and wash plate 5 times with Wash buffer.
9. Add 50 µl of TMB substrate A and 50 µl of TMB substrate B into each well. Gently tap the plate to ensure thorough mixing. Cover the plate and incubate at 37°C in dark for 15 min. And the shades of blue can be seen in the Positive Controls. Negative Controlswells show no obvious color.
10. Add 50 µl of Stop solution into each well and mix thoroughly. The color changes into yellow immediately.

11. Read the O.D. absorbance at 450 nm in a microplate reader immediately after adding the stop solution. (Use the blank well to set zero)(Readings are taken within 30 minutes of termination of reaction)

Data Analysis

Calculation of Results

Negative control: $A \leq 0.1$ (If one well $A > 0.1$, it should be abandoned; If two or three wells $A > 0.1$, it should be retested)

Positive control: $A \geq 0.8$

Calculation of the Cutoff Value

Cutoff Value = $NCx + 0.10$

NCx: Mean Absorbance of Negative Control

Note: Sample with absorbance values \leq Cutoff Value are NON-REACTIVE and are considered NEGATIVE for HV-IgM/IgG.

sample with absorbance values > cutoff value are considered POSITIVE for HV-IgM/IgG.